

Effects of dietary soybean hulls and *Lactobacillus reuteri* on growth performance, nutrient digestibility and noxious gas mission from feces and slurry in finishing pigs

Yan L. , Meng Q. W., Wang J.P. and Kim I. H.

Department of Animal Resource and Science, Dankook University, Choongnam Cheonan, 330-714, Korea

A 6-week experiment was conducted to investigate the effects of supplementation of diets with soybean hulls (NSP) and *Lactobacillus reuteri* (L. reuteri) on growth performance, the apparent total tract digestibility (ATTD) of dry matter (DM) and nitrogen (N) and fecal and slurry noxious gas emission in finishing pigs. A total of 96 pigs with an average initial body weight (BW) of 80.70 ± 2.29 kg were assigned on the basis of weight to four treatment groups (6 replicates per treatment and 4 pigs per pen). In this study, dietary L. reuteri led to a higher ATTD of N than the negative groups ($P < 0.05$). Pigs fed with soybean hull supplemental diet decreased ($P < 0.05$) the fecal NH₃, H₂S and acetic acid emission compared with those without supplementation. The inclusion of L. reuteri significantly decreased ($P < 0.05$) NH₃ emission from slurry as compared with those without supplementation. Moreover, a synergistic effect ($P < 0.05$) between dietary soybean hulls and L. reuteri was observed on the emission of H₂S and R. SH emission from slurry in the current study. Taken together, both the supplementation of soybean hull or L. reuteri could decrease the noxious gas emission without any effect on the animals. Moreover, a synergistic effect between dietary soybean hulls and L. reuteri was observed on the H₂S and R. SH emission in finishing pigs.

Keywords: Soybean hulls/*Lactobacillus reuteri*/Noxious gas emission/Finishing pig

INTRODUCTION

It is well accepted that gas emissions from livestock production systems could result in various environmental problems and should be reduced. Schiffman, (1998) have suggested that pig manure contains several odor producing compounds, including NH₃, H₂S, R.SH and volatile organic compounds. Thus, many studies are being conducted to create new methods to minimize the environmental impacts of meat production because of the increasing pressure to sustain and improve the environment. Generally, soybean hulls are regarded as an inexpensive by-product of soybean processing, which contains high levels of non-starch polysaccharides (NSP). Previous researches have demonstrated that NSP supplementation can reduce nitrogen excretion, noxious gas and volatile fatty acid (VFA) production from manure (Canh et al., 1997; Mroz et al., 2000). DeCam et al., (2001) shown that the addition of non-starch polysaccharides (NSP) has a positive impact on odor concentration in pig houses without hindering growth performance.

It is well suggested that noxious gas emission is highly related to utilization of nutrients and intestinal microbiota ecosystems (Ferket et al. (2002). Elina et al. (2003) have suggested that *Lactobacillus reuteri* are generally safe and are able to survive throughout the gastrointestinal tract. Han et al. (2001) also reported that supplementation with *Lactobacilli* is an effective way to reduce environmental pollutants from animal slurry because it improves feed efficiency and nutrients retention. Therefore, the present study was conducted to investigate the effects of supplementation with soybean hulls and L. reuteri on growth performance, apparent nutrient digestibility and fecal and slurry noxious gas emission in finishing pigs. 2.

MATERIALS AND METHODS

Experimental design, animals, housing and diets

This study was conducted using a 2 (control diet or soybean hulls diet) × 2 (L. reuteri, + or -) factorial experiments. A total of 96 crossbred ([Landrace×Yorkshire]×Duroc) pigs with an average initial BW of 80.70 ± 2.29 kg were used in a 6-week feeding trial to evaluate the effects of dietary soybean hulls and *Lactobacillus reuteri* supplementation on growth performance, nutrient digestibility and noxious gas emission in finishing pigs. At the beginning of the experiment, pigs were allotted to groups on the

basis of their initial BW according to a completely randomized design. There were 6 replicate pens per treatment and 4 pigs per pen (2 gilts and 2 barrows). All diets were fed in meal form and formulated to meet or exceed the NRC (1998) recommendations for all nutrients (Table 1 and 2). Soybean hulls were used as a source of NSP in the diets and L. reuteri was added to the basal diet at the expense of corn. Each pen was equipped with a self-feeder and a nipple waterer to allow ad libitum access to feed and water throughout the experimental period.

Table 1. Diet composition (as fed basis)

Ingredient, %	Control diet	Soybean hulls diet
Corn	70.82	66.98
Soybean meal, dehulled	21.00	19.95
Soybean Hulls	-	5.00
Molasses	4.00	3.80
Tallow	2.00	1.90
Calcium carbonate	1.19	1.19
Difluorinated phosphate	0.46	0.46
NaCl	0.15	0.15
L-Lysine HCl	-	0.15
DL-Methionine	-	0.02
L-threonine	-	0.02
Choline chloride	0.03	0.03
Premix ¹	0.35	0.35

¹Supplied per kg diet: 4,000 IU vitamin A, 800 IU vitamin D₃, 171 IU vitamin E, 2 mg vitamin K, 4 mg vitamin B₂, 1 mg vitamin B₆, 16 g vitamin B₁₂, 11 mg pantothenic acid, 20 mg niacin and 0.08 mg biotin; 15 mg Cu, 100 mg Fe, 100 mg Zn, 40 mg Mn, 0.3 mg I, 0.5 mg Co and 0.4 mg Se.

The probiotic preparation used in current experiment is manufactured by Genebiotech Co., Ltd. (Republic of Korea) under the name of Lactocare®. It is guaranteed to contain at least 1.5×10^6 CFU/g of *Lactobacillus reuteri* GB-LC 1 in a liquid form.

Table 2. Nutrient composition of experimental diets (as fed basis)

Items	Control diet	Soybean hulls diet
Calculated composition		
DE, kcal/kg	3,490	3,420
CP, %	16.00	15.80
CF, %	2.38	3.97
EE, %	2.98	2.94
Ash, %	2.58	2.69
Starch, %	45.40	43.10
Sugar, %	4.95	4.78
Sulfur, %	0.19	0.18
NSP ¹ , %	16.50	19.40
Lys, %	0.78	0.78
Met, %	0.26	0.26
Cys, %	0.29	0.28
Met+Cys, %	0.55	0.54
Trp, %	0.16	0.16
Thr, %	0.60	0.57
Tyr, g/kg	0.57	0.56
Analysis composition, %		
DM	82.50	82.90
CP	16.80	16.20
Ca	0.71	0.70
P	0.60	0.59
ADF	5.31	6.11
NDF	7.40	11.82

1Non-starch polysaccharides (NSP) were determined as DM – Ash – (Crude protein + Crude fat + Starch + Sugar).

Sampling and measurements

Body weight and feed consumption were measured at the end of the experiment to monitor the average daily gain (ADG), average daily feed intake (ADFI) and gain/ feed (G/F) ratio. Chromium oxide (Cr2O3) was added to the diet at 0.20% as an indigestible marker at the beginning of the 6th week to calculate the apparent total tract digestibility for dry matter (DM) and nitrogen (N). Fecal grab samples were then collected randomly from at least two pigs in each pen (1 gilt and 1 barrow) after the pigs were fed diet containing the indicator for 4 d. All feed and fecal samples from one pen were then pooled and mixed, after which the fecal samples were dried for 72 hours at 70 C, and were finely ground to be able to pass through a 1-mm sieve screen and then frozen and stored in a refrigerator at -20 C until analysis. All the feed and fecal samples were then analyzed for DM and N according to the AOAC (1995). Chromium levels were determined via by using UV absorption spectrophotometry (Shimadzu, UV-1201, Japan) and the apparent digestibility of DM and N were calculated using indirect-ratio methods.

Fresh feces and urine samples were collected from two pigs in each pen on the last 2 days of 5th week of the experiment and then mixed well for each respective pen. Analyses were then conducted using fresh fecal samples (300 g) and feces +urine samples (150 g + 150 ml) obtained from each pen that were stored in 2.6-L sealed plastic boxes. Each box had a small hole in the middle of one side wall that was sealed by adhesive plaster and all samples were collected and analyzed in duplicates. After being sealed in the boxes, samples were allowed to ferment for a period of 30 days at room temperature (21 °C) following the method described by Otto et al. (2003). After the fermentation period, the gases produced were evaluated using a Gastec (model GV-100) gas sampling pump (Gastec Corp., Gastec detector tube No. 3L and 3La for NH₃; No. 4LL and 4LK for H₂S; No. 81 and 81L for acetic acid; No. 70 and 70L for R.SH, Gastec Corp, detector tube, Japan). Prior to measurement, the slurry samples were manually shaken for approximately 30 s to disrupt any crust formation on the surface of the slurry sample and to homogenize them. The adhesive plaster was then punctured and 100 mL of the headspace air was sampled approximately 2.0 cm above the feces or slurry surface. Following air sampling, each box was re-sealed with adhesive plaster. The headspace measurement was repeated 24 and 48 h after the initial measurement and the concentration of gases were determined using the average of the 3 head space measurements.

Statistical analysis

The experimental data were analyzed by a 2 × 2 factorial using the GLM Procedure described by the Statistics Analysis Systems Institute (1996). The model evaluated the main effect of soybean hulls, *L. reuteri*, and soybean hulls × *L. reuteri*, with the individual pen serving as the unit. The variability of all of the data was expressed as the standard error (SE) and a P<0.05 was considered to be statistically significant.

RESULTS

Growth performance

No significant differences in the ADG, ADFI and G/F of pigs were observed among treatment groups in this study (Table 3).

Table 3. Effect of supplemental soybean hulls and *Lactobacillus reuteri* on growth performance in finishing pigs¹

Items	Diet				SE ³	
	NSP	CON ²		NSP ²		
		-	+	-		+
<i>L. reuteri</i>						
ADG, g	925	913	911	974	26	
ADFI, g	2,869	2,771	2,777	2,802	35	
gain/feed	0.322	0.330	0.328	0.348	0.01	

1Ninety six pigs with an initial body weight of 80.70 ± 2.29 kg.

2Abbreviation: CON, the control diet; NSP, CON + 5% soybean hulls; -, without

Lactobacillus reuteri 0.2% supplementation; +, with *Lactobacillus reuteri* 0.2% supplementation.

³Pooled standard error.

Apparent total tract digestibility (ATTD) of dry matter (DM) and nitrogen (N)

There were no significant differences in DM digestibility observed among the treatments (Table 4). However, N digestibility was significantly greater when pigs fed diets with a *L. reuteri* (P<0.05) than that in control group.

Table 4. Effect of supplemental soybean hulls and *Lactobacillus reuteri* on apparent nutrient digestibility in finishing pigs¹

Items	Diet				SE ³	
	NSP	CON ²		NSP ²		
		-	+	-		+
<i>L. reuteri</i>						
DM, %	72.75	73.76	72.36	73.04	0.82	
N, % ⁴	71.42	76.89	71.51	73.44	1.16	

1Ninety six pigs with an initial body weight of 80.70 ± 2.29 kg.

2Abbreviation: CON, the control diet; NSP, CON + 5% soybean hulls; -, without *Lactobacillus reuteri* 0.2% supplementation; +, with *Lactobacillus reuteri* 0.2% supplementation.

³Pooled standard error.

⁴*L. reuteri* effect, P<0.05.

Fecal and slurry noxious gas emission

In this study, pig fed the soybean hull supplemental diet decreased (P<0.05) the fecal NH₃, H₂S and acetic acid emission as compared with those without supplementation. the inclusion of *L. reuteri* significantly decreased (P<0.05) slurry NH₃ emission compared with those without supplementation. Moreover, a synergistic effect (P<0.05) between dietary soybean hulls and *L. reuteri* was observed on the slurry H₂S and R. SH emission in the current study.

Table 5. Effect of supplemental soybean hulls and *Lactobacillus reuteri* on noxious gas emission and pH in finishing pigs¹

Items, mg/kg	Diet				SE ³	
	NSP	CON ²		NSP ²		
		-	+	-		+
<i>L. reuteri</i>						
NH ₃						
Feces ⁶	31.5	27.5	26.5	25.0	1.0	
Slurry ⁷	110.0	57.5	90.0	50.0	12.3	
H ₂ S						
Feces ⁶	29.0	15.0	65.0	50.0	13.0	
Slurry ⁸	6.3	3.0	8.0	1.0	1.0	
R.SH						
Feces	27.5	25.0	25.0	22.5	7.0	
Slurry ⁸	17.5	7.0	13.0	5.0	1.8	
Acetic acid						
Feces ⁶	17.6	18.5	20.4	19.6	0.8	
Slurry	21.1	20.8	27.5	26.6	1.1	

1Ninety six pigs with an initial body weight of 80.70 ± 2.29 kg.

2Abbreviation: CON, the control diet; NSP, CON + 5% soybean hulls; -, without *Lactobacillus reuteri* 0.2% supplementation; +, with *Lactobacillus reuteri* 0.2% supplementation.

³Pooled standard error.

⁴Feces (300 g) were kept in a sealed box and fermented for 30 d, a Gastec detector was then used to measure noxious gas emission.

⁵Feces (150 g) + urine (150 mL) were kept in a sealed box and fermented for 30 days, a Gastec detector was then used to measure noxious gas emission.

⁶Soybean hulls effect, P<0.05.

⁷*L. reuteri* effect, P<0.05.

⁸Soybean hulls × *L. reuteri* interaction, P<0.05.

DISCUSSION

Growth performance and apparent total tract digestibility of DM and N

Previously, it is well suggested that dietary NSP could decrease the gastrointestinal emptying rate in pigs because of its higher viscous material (Guérin et al., 2001). Dilger et al. (2004) demonstrated that gastrointestinal

pH may be increased by the buffering capacity of NSP, and reduce the gastric secretion in response to the alkaline environment. Therefore, it is reasonable to hypothesize that dietary soybean hulls (NSP) could decrease the growth performance and nutrient digestibility in the current study. However, the inclusion of soybean hulls did not affect the growth performance and nutrient digestibility compared with the no-supplemental soybean hulls diets, which is in agreement with Radar matrix (1996) and Van Oeckel (1998), who suggested that pig fed soybean hulls supplemental diet (5 or 15%) did not affect the growth performance in growing and finishing pigs, respectively. It is well accepted that fermentation of NSP and dietary fiber takes place in the cecum and colon of monogastric animals and can produce increased amount of VFA, which may be responsible for up to 30% of the maintenance energy needs of growing pigs (Varel and Yen, 1997). They also reported that this amount is higher in finishing pigs due to the greater maturity of cecal function. Therefore, we hypothesized the higher NSP level (soybean hulls) in the diet might result in energy production in the hindgut, which would compensate the lower DE levels in the soybean hulls diet. Furthermore, the CP levels in the current study (16.2% and 16.8%) were higher than the NRC requirement (13.2%). This excess protein may be another reason for the absence of any negative effects in pigs that were provided with the diet containing lower nutrient levels.

Jin et al. (1997) suggested that continuous feeding of direct-fed microbes to livestock could maintain the beneficial intestinal microflora by inducing competitive exclusion of pathogenic bacteria, and by exerting antagonistic activity towards pathogenic bacteria. Our previous study also suggested that dietary *Lactobacillus brevis* improved the growth performance of growing-finishing pigs (Chen et al., 2006 a). Thus, it is reasonable to expect beneficial results could be observed in response to supplementation of probiotics such as *Lactobacillus* species. However, no effect was detected for the growth performance in this study, which is in agreement with Nousiainen and Setälä (1993), who found that the inclusion of *Lactobacillus* did not affect the growth performance of growing-finishing pigs. The reason for the absence is likely to be the more developed digestive system, improved immunity and increased resistance to intestinal disorders as pig become older (80 kg) (Nousiainen and Setälä, 1993). Interestingly, the inclusion of *Lactobacillus reuteri* significantly increased the N digestibility in the current study, which is in agreement with Shon et al. (2005), who suggested that 0.2% *Lactobacillus reuteri* did not affect the DM digestibility, but did increase N digestibility during the finishing phase. Previously, Wenk (2000) have suggested that *Lactobacillus* spp. could stimulate and stabilize the digestion processes by enhancing the population of beneficial micro-organisms and improving microbial enzyme activity. Therefore, the increased gut health and microbial enzyme activity may be responsible for the higher N digestion in this study.

Noxious gas emission

In this study, pig fed the soybean hulls led to a significant depression on the noxious gas emission compared with the no-supplemental diet. It is well suggested that the addition of soybean hulls led to a 0.6% decrease in the level of crude protein and its metabolites, which are precursors for odor production in the gut of animals and in slurry stores (Mackie et al., 1998). Kiene and Hines, (1995) have suggested that variations in the concentrations of sulfuric odorous compounds (H₂S and R.SH.) occur primarily as a result of differences in the sulfur-containing protein of diets and the metabolism of sulfur-containing amino acids (methionine, cystine and cysteine). Wang et al. (2008) also suggested that higher NSP diet could enhance microbial activities within the gut of the animal, and might be responsible for the lower excreta pH and higher slurry VFA concentrations. Our study suggested that soybean hull supplemented diet reduced crude protein concentration compared with the none-supplemented diets. Therefore, it is suitable to hypothesize that the decreased protein levels and the improved intestinal environment may be responsible for the depression effect on the odorous compounds in the current study. Moreover, Ji and Kim (2002) found that addition of complex probiotics (*Lactobacillus acidophilus*, *Bacillus* spp. and *Aspergillus oryzae*) could decrease the NH₃

production of pigs by increasing the nutrient digestibility. Our previous study also suggested that bacillus-based probiotic reduced fecal NH₃-N excretion by improving the intestine environment and nutrient digestibility in finishing pigs (Chen et al., 2006b). Similarly, in this study, the inclusion of dietary probiotic significantly decreased the N digestibility. Therefore, the decreased NH₃ emission from slurry in the current study may have occurred as a result of increased digestibility of nitrogen.

Moreover, a significant interaction between soybean hulls and *L. reuteri* was observed on the H₂S and R.SH emission in the current study. Kiene and Hines, (1995) have suggested that variations in the concentrations of sulfuric odorous compounds (H₂S and R.SH.) occur primarily as a result of differences in the sulfur composition of diets and the metabolism of sulfur-containing amino acids (methionine, cystine and cysteine). It is well accepted that the inclusion of NSP could benefit the populations of Bifidobacteria and Lactobacilli in the large intestine (O'Connell et al., 2005). Our previous study also suggested that dietary probiotic could increase the health of the gastro-intestine tract in the animals (Chen et al., 2006 a,b). Therefore, the reason for the synergistic effect between *L. reuteri* and soybean hulls is likely to be due to their beneficial effect on the intestinal bacteria ecosystem and subsequently its protein digestibility. However, the exact underline mechanism is still unknown. Thus, further research need be conducted to investigate the mechanism responsible for the above effects between soybean hulls and dietary *L. reuteri* supplementation.

In conclusion, both the supplementation of soybean hull and *L. reuteri* could decrease the noxious gas emission without adverse effect on the animals. Moreover, a synergistic effect between dietary soybean hulls and *L. reuteri* was observed on the H₂S and R. SH emission in finishing pigs.

REFERENCES

1. AOAC, 2000. Official methods of analysis, 17th edition. Arlington, VA, USA.
2. Canh, T.T., Verstegen, M.W.A., Aarnink, A.J.A., Schrama, J.W., 1997. Influence of dietary factors on nitrogen partitioning and composition of urine and faeces of fattening pigs. *J. Anim. Sci.* 75, 700-706.
3. Chen, Y.J., Min, B.J., Cho, J.H., Kim, H.J., Yoo, J.S., Kim, I.H., 2006a. Effects of dietary *Lactobacillus brevis* supplementation on growth performance, dry matter and nitrogen digestibilities, blood cell counts and fecal odour emission compounds in growing pigs. *J. Anim. Sci. & Technol (Kor.)*. 48, 503-512.
4. Chen, Y.J., Min, B.J., Cho, J.H., Kwon, O.S., Son, K.S., Kim, H.J., Kim, I.H., 2006b. Effects of dietary Bacillus-based probiotic on growth performance, nutrients digestibility, blood characteristics and fecal noxious gas content in finishing pigs. *Asian-Aust. J. Anim. Sci.* 19, 587-592.
5. DeCam, S.A., Hill, B.E., Hankins, S.L., Bundy, D.C., Powers, W.J., 2001. Effects of soybean hulls in commercial diet on pig performance, slurry composition, and selected air quality parameters in swine facilities. *J. Anim. Sci.* 79 (Suppl. 1), 252 (Abstr.).
6. Ferket, P.R., van Heugten, E., van Kempen, T.A., Angel, R., 2002. Nutritional strategies to reduce environmental emissions from non ruminants. *J. Anim. Sci.* 80 (suppl. 2), 168-182.
7. Guérin, S., Ramonet, Y., Le Cloarec, J., Meunier-Salaün, M.C., Malbert, C.H., 2001. Changes in intragastric meal distribution are better predictors of gastric emptying rate in conscious pigs than are meal viscosity or dietary fibre concentration, *Br. J. Nutr.* 85, 343-350.
8. Han, I.K., Lee, J.H., Piao, X.S., Li, D.F., 2001. Feeding and management system to reduce environmental pollution in swine production: A review. *Asian-Aust. J. Anim. Sci.* 14, 432-444.
9. Ji, F., Kim, S.W., 2002. Reducing odour in swine production: Effect of enzymes and probiotics on ammonia production. *J. Anim. Sci.* 80 (suppl. 1).
10. Jin, L.Z., Ho, Y.W., Abdullah, N., Jalaudin, S., 1997. Probiotics in poultry: modes of action. *World's Poult. Sci. J.* 53, 351-368.
11. Kiene, R.P., Hines, M.E., 1995. Microbial formation of dimethyl

- sulfide in anoxic Sphagnum peat. *Appl. Environ. Microbiol.* 61, 2720-2726.
12. Mackie, R.I., Stroot, P.G., Varel, V.H., 1998. Biochemical identification and biological origin of key odor components in livestock waste. *J. Anim. Sci.* 76, 1331-1342.
 13. Nousiainen, J., Setälä, J., 1993. Lactic acid bacteria as animal probiotics. In: *Lactic acid bacteria*. (Ed. S. Salminen and A. von Wright). Marcel Dekker, Inc., New York. USA.
 14. NRC, 1998. Nutrient requirement of pigs (10th Ed.) National Research Council, Academy Press. Washington, DC.
 15. O'Connell, J.M., Callan, J.J., Sweeney, T., O'Doherty, J.V., 2005. The effect of cereal type and exogenous enzyme supplementation in pig diets on nutrient digestibility, intestinal microflora, volatile fatty acid concentration and slurry ammonia emissions from finisher pigs. *Anim. Sci.* 81, 357-364.
 16. Radar matrix., 1996. Feed Table 28/05/1996, Radar NV, Dorpstraat 4, B-9800 Deinze, Belgium.
 17. Riviere, J., Subtil, J.C., Catroux, G., 1974. Etude de l'évolution physico-chimique et microbiologique du lisier de porcs pendant le stockage anaérobie. *Ann. Agron.* 25, 383-401.
 18. SAS., 1996. SAS user's guide. Release 6.12 edition. SAS Institute. Inc Cary NC. USA.
 19. Schiffman, S.S., 1998. Livestock Odors: Implications for Human Health and Well-Being. *J. Anim. Sci.* 76, 1343-1355.
 20. Shon, K.S., Hong, J.W., Kwon, O.S., Min, B.J., Lee, W.B., Kim, I.H., Park, Y.H., Lee, I.S., 2005. Effects of *Lactobacillus reuteri*-based directed microbial supplementation for growing-finishing Pigs. *Asian-Aust. J. Anim. Sci.* 18, 370-375.
 21. Van Oeckel, M.J., Warnants, N., De Paepe, M., Casteels, M., Boucque, C.V., 1998. Effect of fibre-rich diets on the backfat skatole content of entire male pigs. *Livest. Prod. Sci.* 56, 173-180.
 22. Varel, V.H., Yen, J.T., 1997. Microbial perspective on fiber utilization by swine. *J. Anim. Sci.* 75, 2715-2722.
 23. Wang, Y., Cho, J.H., Chen, Y.J., Yoo, J.S., Kim, H.J., Huang, Y., Shin, S.O., Zhou, T.X., Kim, I.H., 2008. Effect of dietary soyabean hulls and metal-amino acid chelated mineral supplementation on growth performance, nutrient digestibility and noxious gas emission in growing pigs. *J. Anim. Feed Sci.* 17, 171-181.
 24. Wang, Y., Chen, Y.J., Cho, J.H., Yoo, J.S., Huang, Y., Kim, H.J., Shin, S.O., Zhou, T.X., Kim, I.H., 2009. Effect of soybean hulls supplementation to finishing pigs on the emission of noxious gases from slurry. *Anim. Sci. J.* 80, 316-321.
 25. Wenk, C., 2000. Recent advances in animal feed additives such as metabolic modifiers, antimicrobial agents, probiotics, enzymes and highly available minerals. *Asian-Aust. J. Anim. Sci.* 13, 86-95.